

# Package ‘bwimage’

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**Type** Package

**Title** Describe Image Patterns in Natural Structures

**Version** 1.3

**Date** 2020-04-22

**Author** Carlos Biagolini-Jr.

**Maintainer** Carlos Biagolini-Jr.<c.biagolini@gmail.com>

**Depends** stats, utils

**Imports** jpeg, png

**Description** A computational tool to describe patterns in black and white images from natural structures. 'bwimage' implemented functions for exceptionally broad subject. For instance, 'bwimage' provide examples that range from calculation of canopy openness, description of patterns in vertical vegetation structure, to patterns in bird nest structure.

**License** GPL (>= 2)

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## R topics documented:

bwimage-package . . . . .	2
aggregation_index . . . . .	3
altitudinal_profile . . . . .	5
compress . . . . .	6
denseness_column . . . . .	7
denseness_row . . . . .	8
denseness_sample . . . . .	9
denseness_total . . . . .	10
heigh_maximum . . . . .	11
heigh_propotion . . . . .	12
heigh_propotion_test . . . . .	13
hole_columm . . . . .	14
hole_row . . . . .	15

hole_section . . . . .	16
hole_section_data . . . . .	17
image_information . . . . .	18
light_gap . . . . .	19
plot_samples . . . . .	20
stretch . . . . .	21
threshold_color . . . . .	22
threshold_image_list . . . . .	23
topline . . . . .	25

<b>Index</b>	<b>26</b>
--------------	-----------

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bwimage-package	<i>Describe Image Patterns in Natural Structures</i>
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## Description

A computational tool to describe patterns in black and white images from natural structures. 'bwimage' implemented functions for exceptionally broad subject. For instance, 'bwimage' provide examples that range from calculation of canopy openness, description of patterns in vertical vegetation structure, to patterns in bird nest structure.

## Details

The DESCRIPTION file:

```

Package:    bwimage
Type:      Package
Title:     Describe Image Patterns in Natural Structures
Version:   1.3
Date:     2020-04-22
Author:    Carlos Biagolini-Jr.
Maintainer: Carlos Biagolini-Jr.<c.biagolini@gmail.com>
Depends:  stats, utils
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```

Index of help topics:

aggregation_index	Aggregation index calculator
altitudinal_profile	Highest black pixel by sections
bwimage-package	Describe Image Patterns in Natural Structures
compress	Compress square to circle
denseness_column	Denseness in column sections
denseness_row	Denseness in row sections
denseness_sample	Denseness in samples

denseness_total	Denseness for whole image
heigh_maximum	Height of the highest black pixel in the image
heigh_propotion	Cumulative denseness for each line
heigh_propotion_test	Cumulative denseness test
hole_columm	Holes description in columns sections
hole_row	Holes description in row sections
hole_section	Hole finder
hole_section_data	Summary of holes information
image_information	Summary of image information
light_gap	Light gap
plot_samples	Plot samples from denseness_sample
stretch	stretch circle to square
threshold_color	Image to matrix - Single
threshold_image_list	Image to matrix - List
topline	Top line

A computational tool to describe patterns in black and white images from natural structures.

#### Author(s)

Carlos Biagolini-Jr.

Maintainer: Carlos Biagolini-Jr.<c.biagolini@gmail.com>

#### References

Biagolini-Jr C, Macedo RH (2019) bwimage: A package to describe image patterns in natural structures. F1000Research 8 Lambers M (2016) Mappings between sphere, disc, and square. Journal of Computer Graphics Techniques Vol 5:1-21 Nobis M, Hunziker U (2005) Automatic thresholding for hemispherical canopy-photographs based on edge detection. Agricultural and forest meteorology 128:243-250 Shirley P, Chiu K (1997) A low distortion map between disk and square. Journal of graphics tools 2:45-52 Zehm A, Nobis M, Schwabe A (2003) Multiparameter analysis of vertical vegetation structure based on digital image processing. Flora-Morphology, Distribution, Functional Ecology of Plants 198:142-160

#### Examples

```
bush<-system.file("extdata/bush.JPG", package ="bwimage")
bush_imagematrix<-threshold_color(bush, "jpeg", "proportional",compress_rate = 0.1)
aggregation_index(bush_imagematrix)
```

## Description

The function `aggregation_index` calculate the aggregation index. It works for matrix with and without transparent pixel. The aggregation index is a standardized estimation of the average proportion of same-color pixels around each image pixel. First, the proportion of same-color neighboring pixels (SCNP) is calculated (marginal lines and columns are excluded). Next, the SCNP for all pixels are averaged; then, given the proportion of black and white pixels, number of pixels in height and width, and location of transparent pixels (when present), the maximum and minimum possible aggregation indexes are calculated. Finally, the observed aggregation is standardized to a scale where the minimum possible value is set at zero and the maximum value is set at one.

## Usage

```
aggregation_index(imagematrix)
```

## Arguments

`imagematrix`      The matrix to be analysed.

## Value

`adjusted_aggregation`  
Standardized aggregation.

`non_adjusted_aggregation`  
Observed aggregation.

## Author(s)

Carlos Biagolini-Jr.

## See Also

`threshold_color`

## Examples

```
# First, get a matrix from your image. Here an example of a bush image is used.
# Using aggregation_index to estimate vegetation agregation
bush<-system.file("extdata/bush.JPG", package ="bwimage")
bush_imagematrix<-threshold_color(bush, "jpeg", "proportional", compress_rate = 0.1)
aggregation_index(bush_imagematrix)

# Using aggregation_index to estimate aggregation of nest wall holes
nestwall<-system.file("extdata/bird_nestwall.png", package ="bwimage")
nestwall_imagematrix<-threshold_color(nestwall, "png", "width_fixed", target_width=300)
aggregation_index(nestwall_imagematrix)
```

---

altitudinal\_profile    *Highest black pixel by sections*

---

### Description

Break the original matrix in a number of section ( n\_sections), then find the higher black pixel in each image section.

### Usage

```
altitudinal_profile(imagematrix, n_sections, height_size)
```

### Arguments

imagematrix	The matrix to be analysed.
n_sections	Break the image in this number of columns.
height_size	Real size of image height (in mm, cm, m, etc..).

### Value

Mean	Height mean of the highest black pixel in sections.
SD	Standard deviations of the highest black pixel in sections.
Size	Height of the highest black pixel in sections.

### Author(s)

Carlos Biagolini-Jr.

### References

Zehm et al 2003 Multiparameter analysis of vertical vegetation structure based on digital image processing. *Flora-Morphology, Distribution, Functional Ecology of Plants*, 198: 142-160.

### See Also

threshold\_color

### Examples

```
# First, get a matrix from your image. Here an example of a bush image is used.
bush<-system.file("extdata/bush.JPG",package ="bwimage")
bush_imagematrix<-threshold_color(bush, "jpeg", "proportional", compress_rate = 0.1)

# Profile of highest black pixels on sections of the bush image matrix
altitudinal_profile(bush_imagematrix,n_sections = 10, height_size=100)
# Conclusions:
# i) the mean height of the highest black pixel is 45.28 cm.
# ii) standard deviation of highest black height is 21.54.
```

compress

*Compress square to circle*

---

**Description**

Compress data from square image to circular in binary matrix

**Usage**

```
compress(imagematrix, method = "radial", background = NA)
```

**Arguments**

<code>imagematrix</code>	The matrix to be compressed.
<code>method</code>	Compress algorithm. Four algorithms (radial, shirley, squircle, and elliptical) are available to stretch the image. The algorithms were adapted from Lambers 2016.
<code>background</code>	Code for background cell value. When compressing a squared matrix, corners of the transformed matrix will no have corresponding pixel from original matrix. Thus, the background value will be the value of transformed matrix corners.

**Value**

A matrix of 0, 1 and NA representing white, black and transparent pixels, respectively.

**Author(s)**

Carlos Biagolini-Jr.

**References**

Lambers 2016 Mappings between Sphere, Disc, and Square. *Journal of Computer Graphics Techniques*, 5(2): 1-21.

**Examples**

```
img_location <- system.file("extdata/chesstable.png", package = "bwimage")
image_matrix <- threshold_color(img_location, "png", "frame_fixed", target_width = 50, target_height = 50)
compress(image_matrix, method = "radial")
```

---

denseness\_column      *Denseness in column sections*

---

### Description

Calculate the denseness (proportion of black pixel in relation to the total number of pixels) for a given number of sections (`n_sections`). `n_sections` should be set as a number, in this situation `denseness_column` will break the original matrix in slices, and apply `denseness_total` function for each section. For instance, in a matrix of 1000x1000 if `n_sections` = 10, it will break to 10 sections of 1000x100 and analyze it. In other words, the sections will be the following sections of the original matrix [1:1000, 1:100] ,[ 1:1000,101:200] ,[ 1:1000,201:300] ,[ 1:1000,301:400] ,[ 1:1000,401:500] ,[ 1:1000,501:600] ,[ 1:1000,601:700] ,[ 1:1000,701:800] ,[ 1:1000,801:900] ,[ 1:1000,901:1000]. The default for parameter `n_sections` is "all", it will calculate denseness for each column of pixel. In other words, it will break the image in a number of section equal to the image pixel width.

### Usage

```
denseness_column(imagematrix, n_sections = "all")
```

### Arguments

<code>imagematrix</code>	The matrix to be analysed.
<code>n_sections</code>	Break the image in this number of columns.

### Value

Denseness	Denseness of each column section.
Mean	Mean of column sections denseness.
SD	standard deviations of column sections denseness.

### Author(s)

Carlos Biagolini-Jr.

### References

Zehm et al 2003 Multiparameter analysis of vertical vegetation structure based on digital image processing. *Flora-Morphology, Distribution, Functional Ecology of Plants*, 198: 142-160.

### See Also

`denseness_total` `threshold_color`

## Examples

```
# First, get a matrix from your image. Here an example of a bush image is used.
bush<-system.file("extdata/bush.JPG",package ="bwimage")
bush_imagematrix<-threshold_color(bush, "jpeg", "proportional", compress_rate = 0.1)

# Calculate vegetation denseness in 20 column sections
denseness_column(bush_imagematrix,20)
```

---

`denseness_row`

*Denseness in row sections*

---

## Description

Calculate the denseness (proportion of black pixel in relation to the total number of pixels) for a given number of sections (`n_sections`). `n_sections` should be set as a number, in this situation `denseness_row` will break the original matrix in slices, and apply `denseness_total` function for each section. For instance, in a matrix of 1000x1000 if `n_sections = 10`, it will break to 10 sections of 100x1000 and analyze it. In other words, the sections will be the following sections of the original matrix [1:100, 1:1000] , [101:200, 1:1000] , [201:300, 1:1000] , [301:400, 1:1000] , [401:500, 1:1000] , [501:600, 1:1000] , [601:700, 1:1000] , [701:800, 1:1000] , [801:900, 1:1000] , [901:1000, 1:1000] .The default for parameter `n_sections` is "all", it will calculate denseness for each row of pixel. In other words, it will break the image in a number of section equal to the image pixel height.

## Usage

```
denseness_row(imagematrix, n_sections = "all")
```

## Arguments

<code>imagematrix</code>	The matrix to be analysed.
<code>n_sections</code>	Break the image in this number of rows.

## Value

Denseness	Denseness of each row section.
Mean	Mean of row sections denseness.
SD	standard deviations of row sections denseness.

## Author(s)

Carlos Biagolini-Jr.

## References

Zehm et al 2003 Multiparameter analysis of vertical vegetation structure based on digital image processing. *Flora-Morphology, Distribution, Functional Ecology of Plants*, 198: 142-160.



**See Also**

denseness\_total threshold\_color

**Examples**

```
# First, get a matrix from your image. Here an example of a bush image is used.
bush<-system.file("extdata/bush.JPG",package ="bwimage")
bush_imagematrix<-threshold_color(bush, "jpeg", "proportional",compress_rate = 0.1)

# Calculate vegetation denseness in 20 row sections
denseness_row(bush_imagematrix, n_sections = 20)
```

---

denseness\_sample      *Denseness in samples*

---

**Description**

Calculate the denseness (proportion of black pixel in relation to the total number of pixels) for a given number of samples.

**Usage**

```
denseness_sample(imagematrix, width_size, height_size, sample_width,
  sample_height, method = "random", sample_shape = "rectangle",
  n_samples = 10, n_sample_horizontal = 10, n_sample_vertical = 1,
  proportion_horizontal = 1, proportion_vertical = 1,
  align_horizontal = "center", align_vertical = "bottom")
```

**Arguments**

imagematrix	The matrix to be analysed.
width_size	Real size of image width (in mm, cm, m, etc..).
height_size	Real size of image height (in mm, cm, m, etc..).
sample_width	Width of sample area.
sample_height	Height of sample area.
method	Method for sample ("random" or "uniform").
sample_shape	The shape of sample unity ("rectangle" or "ellipse"). See plot_samples function.
n_samples	Defines the number of samples, when sample_shape="random".
n_sample_horizontal	Defines the number of samples column, when sample_shape=" uniform".
n_sample_vertical	Defines the number of samples lines, when sample_shape=" uniform".
proportion_horizontal	Range from 0 to 1. Represent the proportion of horizontal plane to be sample. If proportion_horizontal=1 (default) all columns beacome potentially sample.

`proportion_vertical`  
 Range from 0 to 1. Represent the proportion of vertical plane to be sample. If `proportion_vertical=1` (default) all lines become potentially sample.

`align_horizontal`  
 Define horizontal align. Three options are available: "center", "left" or "right".

`align_vertical`  
 Define vertical align. Three options are available: "middle", "bottom" or "top".

**Value**

`Sample_denseness`  
 Proportion of black pixels in samples. It do not take into account transparent pixels (when present)..

`Height`  
 Height of each sample (in mm, cm, m, etc. ..). Central point used as reference.

`Distance(left)`  
 Distance ti the left side of each sample (in mm, cm, m, etc. ..). Central point used as reference.

`Matrix(line)`  
 Imagem matrix line coordinates.

`Matrix(column)`  
 Imagem matrix column coordinates.

**Author(s)**

Carlos Biagolini-Jr.

**See Also**

`plot_samples`

**Examples**

```

# Get a matrix from your image. Here examples provided by bwimage package.

bush<-system.file("extdata/bush.JPG",package ="bwimage")
bush<-threshold_color(bush, "jpeg", "proportional",compress_rate = 0.1)
denseness_sample(bush, width_size=100, height_size=100, sample_width=5, sample_height=5)

```

---

`denseness_total`      *Denseness for whole image*

---

**Description**

Proportion of black pixels in relation to all pixels. It do not take into account transparent pixels (when present).

**Usage**

```
denseness_total(imagematrix)
```

**Arguments**

imagematrix      The matrix to be analysed.

**Value**

Proportion of black pixels in relation to all pixels. It do not take into account transparent pixels (when present).

**Author(s)**

Carlos Biagolini-Jr.

**References**

Zehm et al 2003 Multiparameter analysis of vertical vegetation structure based on digital image processing. *Flora-Morphology, Distribution, Functional Ecology of Plants*, 198: 142-160.

**See Also**

threshold\_color

**Examples**

```
# Get a matrix from your image. Here examples provided by bwimage package.

# I) Calculate vegetation denseness
bush<-system.file("extdata/bush.JPG",package ="bwimage")
bush_imagematrix<-threshold_color(bush, "jpeg", "proportional",compress_rate = 0.1)
denseness_total(bush_imagematrix)

# II) Calculate canopy openness
# Convert image into binary matrix
canopy<-system.file("extdata/canopy.JPG",package ="bwimage")
canopy_matrix<-threshold_color(canopy,"jpeg", compress_method="proportional",compress_rate=0.1)
1-denseness_total(canopy_matrix) # canopy openness
```

---

heigh\_maximum      *Height of the highest black pixel in the image*

---

**Description**

Find the higher black pixel in the whole image.

**Usage**

```
heigh_maximum(imagematrix, height_size)
```

**Arguments**

`imagematrix`     The matrix to be analysed.  
`height_size`     Real size of image width (in mm, cm, m, etc..).

**Value**

Height of the highest black pixel. It is scaleted for the real size (in mm, cm, m, etc..) based in the information from argument `height_size`.

**Author(s)**

Carlos Biagolini-Jr.

**References**

Zehm et al 2003 Multiparameter analysis of vertical vegetation structure based on digital image processing. *Flora-Morphology, Distribution, Functional Ecology of Plants*, 198: 142-160.

**See Also**

`threshold_color`

**Examples**

```

# First, get a matrix from your image. Here an example of a bush image is used.
bush<-system.file("extdata/bush.JPG",package ="bwimage")
bush_imagematrix<-threshold_color(bush, "jpeg", "proportional", compress_rate = 0.1)

# Calculate height of the highest black pixel in the bush image matrix
heigh_maximum(bush_imagematrix,height_size=100)
# Conclusions: The highest vegetation unit ,i.e. highest black pixel, is 84.4 cm above ground.
  
```

---

`heigh_propotion`     *Cumulative denseness for each line*

---

**Description**

Proportion of black pixel below each matrix line.

**Usage**

```
heigh_propotion(imagematrix)
```

**Arguments**

`imagematrix`     The matrix to be analysed.

**Author(s)**

Carlos Biagolini-Jr.

**References**

Zehm et al 2003 Multiparameter analysis of vertical vegetation structure based on digital image processing. *Flora-Morphology, Distribution, Functional Ecology of Plants*, 198: 142-160.

**See Also**

threshold\_color

**Examples**

```
# First, get a matrix from your image. Here an example of a bush image is used.
bush<-system.file("extdata/bush.JPG",package ="bwimage")
bush_imagematrix<-threshold_color(bush, "jpeg", "proportional", compress_rate = 0.1)

# Proportion of black pixel below each matrix line.
heigh_propotion(bush_imagematrix)
```

---

heigh\_propotion\_test *Cumulative denseness test*

---

**Description**

Find the height which a given proportion of black pixel is found.

**Usage**

```
heigh_propotion_test(imagematrix, proportion, height_size)
```

**Arguments**

imagematrix	The matrix to be analysed.
proportion	Proportion of denseness to test.
height_size	Real size of image height (in mm, cm, m, etc..).

**Author(s)**

Carlos Biagolini-Jr.

**References**

Zehm et al 2003 Multiparameter analysis of vertical vegetation structure based on digital image processing. *Flora-Morphology, Distribution, Functional Ecology of Plants*, 198: 142-160.

**See Also**

threshold\_color

**Examples**

```
# First, get a matrix from your image. Here an example of a bush image is used.
bush<-system.file("extdata/bush.JPG",package ="bwimage")
bush_imagematrix<-threshold_color(bush, "jpeg", "proportional", compress_rate = 0.1)

# See the proportion of black pixels (1) below each bush image matrix row
heigh_propotion_test(bush_imagematrix,0.75,100)
# Conclusion: in this imagem, 75 percent of the vegetation is hold below 31.2 cm.
```

---

hole\_columm

*Holes description in columns sections*

---

**Description**

Summary information of holes in a given number of columns (`n_sections`). `n_sections` must be set as a number, in this situation `hole_columm` will sample columns, and apply `hole_section_data` function for each section. Next, all results will be display on `hole_columm` output. Example of how column sample works: in a matrix of 250x250 if `n_sections = 5` , it will sample columns 1,51,101,151, and 201 and analyze it. In other words, the sections will be following sections of the original matrix [1:250,1] , [1:250,51], [1:250,101], [1:250,151], [1:250,201]. The default for parameter `n_sections` is "all", it will calculate `hole_section_data` for each column of pixel. In other words, it will break the image in a number of section equal to the image pixel width.

**Usage**

```
hole_columm(imagematrix, color = 0, n_sections = "all")
```

**Arguments**

<code>imagematrix</code>	The matrix to be analysed.
<code>color</code>	Color of the hole (0 or 1).
<code>n_sections</code>	Sample this number of columns.

**Value**

N	Number of sections.
Mean	Mean sections size.
SD	standard deviations of sections size.
Min	Minimum sections size sections size.
Max	Maximum sections size.
LH	Stratum with largest hole count.

**Author(s)**

Carlos Biagolini-Jr.

**References**

Zehm et al 2003 Multiparameter analysis of vertical vegetation structure based on digital image processing. *Flora-Morphology, Distribution, Functional Ecology of Plants*, 198: 142-160.

**See Also**

hole\_section\_data threshold\_color

**Examples**

```
# First, get a matrix from your image. Here an example of a bush image is used.
bush<-system.file("extdata/bush.JPG",package ="bwimage")
bush_imagematrix<-threshold_color(bush, "jpeg", "proportional", compress_rate = 0.1)

# Information of white (i.e. 0s in matrix) holes in 5 columns uniformly sample among matrix.
hole_columm(bush_imagematrix, n_sections=5 )

# Information of black (i.e. 1s in matrix) holes in 20 columns uniformly sample among matrix.
hole_columm(bush_imagematrix, n_sections=20 )
```

---

hole\_row

*Holes description in row sections*

---

**Description**

Summary information of holes in a given number of rows (`n_sections`). `n_sections` must be set as a number, in this situation `hole_row` will sample rows, and apply `hole_section_data` function for each section. Next, all results will be display on `hole_columm` output. Example of how row sample works: in a matrix of 250x250 if `n_sections = 5` , it will sample rows 1,51,101,151, and 201 and analyze it. In other words, the sections will be following sections of the original matrix [1,1:250] , [51,1:250] , [101,1:250] , [151,1:250] , [201,1:250]. The default for parameter `n_sections` is "all", it will calculate `hole_section_data` for each row of pixel. In other words, it will break the image in a number of section equal to the image pixel height.

**Usage**

```
hole_row(imagematrix, color = 0, n_sections = "all")
```

**Arguments**

<code>imagematrix</code>	The matrix to be analysed.
<code>color</code>	Color of the hole (0 or 1).
<code>n_sections</code>	Sample this number of rows.

**Value**

N	Number of sections.
Mean	Mean sections size.
SD	standard deviations of sections size.
Min	Minimum sections size sections size.
Max	Maximum sections size.
LH	Stratum with largest hole count.

**Author(s)**

Carlos Biagolini-Jr.

**See Also**

hole\_section\_data threshold\_color

**Examples**

```
# First, get a matrix from your image. Here an example of a bush image is used.
bush<-system.file("extdata/bush.JPG",package ="bwimage")
bush_imagematrix<-threshold_color(bush, "jpeg", "proportional", compress_rate = 0.1)

# Information of white (i.e. 0s in matrix) holes in 10 rows uniformly sample among matrix.
hole_row(bush_imagematrix, n_sections=10)

# Information of black (i.e. 1s in matrix) holes in 15 rows uniformly sample among matrix.
hole_row(bush_imagematrix, n_sections=15)
```

---

hole\_section

*Hole finder*

---

**Description**

Description of when a sequence of same color pixel start and end.

**Usage**

```
hole_section(section)
```

**Arguments**

section            Section to be analysed.

**Value**

Description of start and end of each same color sequence



**Author(s)**

Carlos Biagolini-Jr.

**References**

Zehm et al 2003 Multiparameter analysis of vertical vegetation structure based on digital image processing. *Flora-Morphology, Distribution, Functional Ecology of Plants*, 198: 142-160.

**See Also**

hole\_section\_data threshold\_color

**Examples**

```
# First, get a matrix from your image. Here an example of a bush image is used.
bush<-system.file("extdata/bush.JPG",package ="bwimage")
bush_imagematrix<-threshold_color(bush, "jpeg", "proportional", compress_rate = 0.1)

# Find pixel hole sections in the column 200 of bush image
hole_section(bush_imagematrix[,200])

# Find pixel hole sections in the row 250 of bush image
hole_section(bush_imagematrix[250,])
```

---

hole\_section\_data      *Summary of holes information*

---

**Description**

Summary information of holes of a given color in a given section. Result unit is the number of cell.

**Usage**

```
hole_section_data(section, color = 0)
```

**Arguments**

section	Section to be analysed.
color	Color of the hole (0 or 1).

**Value**

N	Number of hole sections
Mean	Mean size of hole sections
SD	Standard deviation of hole sections size
Min	Minimum size of hole sections
Max	Maximum size of hole sections

**Author(s)**

Carlos Biagolini-Jr.

**References**

Zehm et al 2003 Multiparameter analysis of vertical vegetation structure based on digital image processing. *Flora-Morphology, Distribution, Functional Ecology of Plants*, 198: 142-160.

**See Also**

hole\_section threshold\_color

**Examples**

```
# First, get a matrix from your image. Here an example of a bush image is used.
bush<-system.file("extdata/bush.JPG",package ="bwimage")
bush_imagematrix<-threshold_color(bush, "jpeg", "proportional", compress_rate = 0.1)

# Detail information of white (0) holes sections in the column 200 of bush image
hole_section_data(bush_imagematrix[,200], color = 0)

# Detail information of black (1) holes sections in the row 250 of bush image
hole_section_data(bush_imagematrix[250,], color = 1)
```

---

image\_information      *Summary of image information*

---

**Description**

Provide the information of: number of black, white and transparent pixels, total number of pixels, height and width size.

**Usage**

```
image_information(imagematrix)
```

**Arguments**

imagematrix      The matrix to be analysed.

**Value**

Black	Number of black pixels
White	Number of white pixels
Transparent	Number of transparent pixels
Total	Total number of pixels
Height	Size in height
Width	Size in width

**Author(s)**

Carlos Biagolini-Jr.

**See Also**

threshold\_color

**Examples**

```
# First, get a matrix from your image. Here an example of a bush image is used.
bush<-system.file("extdata/bush.JPG",package ="bwimage")
bush_imagematrix<-threshold_color(bush, "jpeg", "proportional",compress_rate = 0.1)
image_information(bush_imagematrix)
```

---

light\_gap

*Light gap*

---

**Description**

Left and right distances from first black pixel to image edge.

**Usage**

```
light_gap(imagematrix, width_size = NA, scale = TRUE)
```

**Arguments**

imagematrix	The matrix to be analysed
width_size	Real size of image width (in mm, cm, m, etc..).
scale	If FALSE do not ajust the output for real size.

**Value**

Distances without black pixel in each side of the picture

**Author(s)**

Carlos Biagolini-Jr.

**References**

Zehm et al 2003 Multiparameter analysis of vertical vegetation structure based on digital image processing. *Flora-Morphology, Distribution, Functional Ecology of Plants*, 198: 142-160.

**See Also**

threshold\_color

**Examples**

```
# First, get a matrix from your image. Here an example of a bush image is used.
bush<-system.file("extdata/bush.JPG",package ="bwimage")
bush_imagematrix<-threshold_color(bush, "jpeg", "proportional", compress_rate = 0.1)

# Calculate vegetation Light gap in the bush image matrix
light_gap(bush_imagematrix,width_size=100)
# Conclusion: there is no light gap on both sides of bush image.
```

---

plot\_samples

*Plot samples from denseness\_sample*


---

**Description**

Plot samples from denseness\_sample.

**Usage**

```
plot_samples(imagematrix, central_lines, central_collumns, width_size,
             height_size, sample_width, sample_height, sample_shape)
```

**Arguments**

imagematrix	The matrix to be analysed.
central_lines	Lines data (i.e. "Matrix(line)") provided by denseness_sample
central_collumns	Collumns data (i.e. "Matrix(column)") provided by denseness_sample
width_size	Real size of image width (in mm, cm, m, etc..).
height_size	Real size of image height (in mm, cm, m, etc..).
sample_width	Width of sample area.
sample_height	Height of sample area.
sample_shape	Inform the shape of sample unity used ("rectangle" or "ellipse"). See denseness_sample function.

**Value**

Plot of the analysed matrix (black and white) and sample locations (red).

**Author(s)**

Carlos Biagolini-Jr.

**See Also**

denseness\_sample

## Examples

```
# Get a matrix from your image. Here examples provided by bwimage package.

bush<-system.file("extdata/bush.JPG",package ="bwimage")
bush<-threshold_color(bush, "jpeg", "proportional",compress_rate = 0.1)
a<-denseness_sample(bush, width_size=100, height_size=100, sample_width=5, sample_height=5)
plot_samples(bush, a[,4],a[,5], 100,100, 5, 5,"rectangle")
```

---

stretch	<i>stretch circle to square</i>
---------	---------------------------------

---

## Description

Stretch data from circular image to square in binary matrix

## Usage

```
stretch(imagematrix, method = "radial")
```

## Arguments

imagematrix	The matrix to be stretched.
method	Stretch algorithm. Four algorithms (radial, shirley, squircle, and elliptical) are available to stretch the image. The algorithms were adapted from Lambers 2016.

## Value

A matrix of 0, 1 and NA representing white, black and transparent pixels, respectively.

## Author(s)

Carlos Biagolini-Jr.

## References

Lambers 2016 Mappings between Sphere, Disc, and Square. *Journal of Computer Graphics Techniques*, 5(2): 1-21.

## Examples

```
img_location <- system.file("extdata/chesstable.png",package ="bwimage")
image_matrix<- threshold_color(img_location,"png", "frame_fixed",target_width = 50,target_height=50)
stretch(image_matrix,method="radial")
```

---

threshold_color	<i>Image to matrix - Single</i>
-----------------	---------------------------------

---

### Description

Convert a single image into a matrix

### Usage

```
threshold_color(filename, filetype = "jpeg", compress_method = "none",
               compress_rate = 1, target_width = 100, target_height = 100,
               threshold_value = 0.5, transparency_regulation = 0.5,
               channel = "rgb")
```

### Arguments

filename	Name of the file to be load - ex: "Figure01.JPG".
filetype	Type of the file to be load. Compatible file types: ".JPGE", ".JPG" or ".PNG".
compress_method	For high resolution files, i.e. numbers of pixels in width and height, it is suggested to reduce the resolution to create a smaller matrix, it strongly reduce GPU usage and time necessary to run analyses. On the other hand, by reducing resolution, it will also reduce the accuracy of data description. The available methods for image reduction are: i) frame_fixed, which resamples images to a desired target width and height; ii) proportional, which resamples the image by a given ratio provided in the argument "proportion"; iii) width_fixed, which resamples images to a target width, and also reduces the image height by the same factor. For instance, if the original file had 1000 pixels in width, and the new width_was set to 100, height will be reduced by a factor of 0.1 (100/1000); and iv) height_fixed, analogous to width_fixed, but assumes height as reference.
compress_rate	Compress rate to by apply if compress_method=proportional. Note: it should be ser as number range from 0 to 1 .
target_width	Target width to be used if compress_method=frame_fixed or compress_method=width_fixed.
target_height	Target height to be used if compress_method=frame_fixed or compress_method=height_fixed.
threshold_value	For each pixel, the intensity of color channels (red, green and blue) are averaged and compared to a threshold_value (threshold). If the average intensity is less than the threshold_value (default is 0.5) the pixel will be set as black, otherwise it will be white. See channel argument.
transparency_regulation	For PNG images, the alpha channel is used to set transparent pixels, i.e. alpha channel values above transparency_regulation (a threshold) will set the pixel as transparent, default is 0.5. NOTE: In the data matrix the value 1 represents black pixels, 0 represents white pixels and NA represents transparent pixels.

**channel** RGB channel to be considered in threshold. If channel=RGB (default), the intensity of red, green and blue is averaged and compared to threshold\_value. If the average intensity is less than the threshold\_value (default is 50 If only one channel is defined ("R" for red, "G" for green, and "B" for blue), the average intensity selected channel compared direct to the threshold\_value value.

### Value

A matrix of 0, 1 and NA representing white, black and transparent pixels, respectively.

### Author(s)

Carlos Biagolini-Jr.

### Examples

```
bush<-system.file("extdata/bush.JPG", package ="bwimage")
threshold_color(bush,"jpeg", "frame_fixed",target_width = 15,target_height=15)

# For your images, if the file is in the working directory type:
# threshold_color("FILE_NAME.EXTENSION", filetype ="FILE_EXTENSION")
# or, if the file is in the other directory:
# threshold_color("C:/PATH TO FILE FOLDER/YOUR_FILE_NAME.EXTENSION", "FILE_EXTENSION")
```

---

threshold\_image\_list *Image to matrix - List*

---

### Description

Convert two or more images into a list of matrices

### Usage

```
threshold_image_list(list_names, filetype = "jpeg",
  compress_method = "none", compress_rate = 1, target_width = 100,
  target_height = 100, threshold_value = 0.5,
  transparency_regulation = 0.5, channel = "rgb")
```

### Arguments

**list\_names** An object contains the names of the files.

**filetype** Type of the file to be load. Compatible file types: ".JPGE", ".JPG" or ".PNG".

**compress\_method**

For high resolution files, i.e. numbers of pixels in width and height, it is suggested to reduce the resolution to create a smaller matrix, it strongly reduce GPU usage and time necessary to run analyses. On the other hand, by reducing resolution, it will also reduce the accuracy of data description. The available methods for image reduction are: i) `frame_fixed`, which resamples images to a

	desired target width and height; ii) proportional, which resamples the image by a given ratio provided in the argument "proportion"; iii) width_fixed, which resamples images to a target width, and also reduces the image height by the same factor. For instance, if the original file had 1000 pixels in width, and the new width_was set to 100, height will be reduced by a factor of 0.1 (100/1000); and iv) height_fixed, analogous to width_fixed, but assumes height as reference.
compress_rate	Compress rate to be applied if compress_method=proportional. Note: it should be set as a number ranging from 0 to 1.
target_width	Target width to be used if compress_method=frame_fixed or compress_method=width_fixed.
target_height	Target height to be used if compress_method=frame_fixed or compress_method=height_fixed.
threshold_value	For each pixel, the intensity of color channels (red, green and blue) are averaged and compared to a threshold_value (threshold). If the average intensity is less than the threshold_value (default is 0.5) the pixel will be set as black, otherwise it will be white. See channel argument.
transparency_regulation	For PNG images, the alpha channel is used to set transparent pixels, i.e. alpha channel values above transparency_regulation (a threshold) will set the pixel as transparent, default is 0.5. NOTE: In the data matrix the value 1 represents black pixels, 0 represents white pixels and NA represents transparent pixels.
channel	RGB channel to be considered in threshold. If channel=RGB (default), the intensity of red, green and blue is averaged and compared to threshold_value. If the average intensity is less than the threshold_value (default is 50) if only one channel is defined ("R" for red, "G" for green, and "B" for blue), the average intensity of the selected channel is compared directly to the threshold_value.

**Value**

A matrix of 0, 1 and NA representing white, black and transparent pixels, respectively.

**Author(s)**

Carlos Biagolini-Jr.

**See Also**

threshold\_color

**Examples**

```
# Image examples provided by bwimage package
bush<-system.file("extdata/bush.JPG", package = "bwimage")
canopy<-system.file("extdata/canopy.JPG", package = "bwimage")

# Convert images to a list of matrices
working_matrices<-threshold_image_list(c(bush,canopy), "jpeg", "proportional", compress_rate = 0.1)
```



---

topline	<i>Top line</i>
---------	-----------------

---

**Description**

Line running along the crest of highest black pixel.

**Usage**

```
topline(imagematrix, height_size = NA, width_size = NA)
```

**Arguments**

<code>imagematrix</code>	The matrix to be analysed.
<code>height_size</code>	Real size of image height (in mm, cm, m, etc..).
<code>width_size</code>	Real size of image width (in mm, cm, m, etc..).

**Value**

Top line size that cover black pixels

**Author(s)**

Carlos Biagolini-Jr.

**References**

Zehm et al 2003 Multiparameter analysis of vertical vegetation structure based on digital image processing. *Flora-Morphology, Distribution, Functional Ecology of Plants*, 198: 142-160.

**See Also**

`threshold_color`

**Examples**

```
# First, get a matrix from your image. Here an example of a bush image is used.
bush<-system.file("extdata/bush.JPG",package = "bwimage")
bush_imagematrix<-threshold_color(bush, "jpeg", "proportional", compress_rate = 0.1)

# See the proportion of black pixels (1) below each bush image matrix row
topline(bush_imagematrix,100,100)
# Conclusion: topline size is 785.6 cm.
```

# Index

- \* **animal**
  - [bwimage-package, 2](#)
- \* **ecology methods**
  - [bwimage-package, 2](#)
- \* **field**
  - [bwimage-package, 2](#)
- \* **image analyses**
  - [bwimage-package, 2](#)
- \* **image processing**
  - [bwimage-package, 2](#)
- \* **vegetation patterns**
  - [bwimage-package, 2](#)

[aggregation\\_index, 3](#)  
[altitudinal\\_profile, 5](#)

[bwimage \(bwimage-package\), 2](#)  
[bwimage-package, 2](#)

[compress, 6](#)

[denseness\\_column, 7](#)  
[denseness\\_row, 8](#)  
[denseness\\_sample, 9](#)  
[denseness\\_total, 10](#)

[heigh\\_maximum, 11](#)  
[heigh\\_propotion, 12](#)  
[heigh\\_propotion\\_test, 13](#)  
[hole\\_column, 14](#)  
[hole\\_row, 15](#)  
[hole\\_section, 16](#)  
[hole\\_section\\_data, 17](#)

[image\\_information, 18](#)

[light\\_gap, 19](#)

[plot\\_samples, 20](#)

[stretch, 21](#)

[threshold\\_color, 22](#)  
[threshold\\_image\\_list, 23](#)  
[topline, 25](#)